

## PROXIMATE COMPOSITION, FUNCTIONAL AND SENSORY PROPERTIES OF FERMENTED MAIZE (*ZEA MAYS*) AND MORINGA (*MORINGA OLEIFERA*) SEED PROTEIN CONCENTRATE FLOUR BLENDS

Olufunke Oluseyi **Ezekiel**<sup>1</sup> and Olajide Emmanuel **Adedeji**<sup>2\*</sup>

<sup>1</sup>Department of Food Technology, University of Ibadan, Ibadan, Nigeria.

<sup>2</sup>Department of Food Science and Technology, Federal University Wukari, PMB 1020, Wukari, Nigeria.

\*Corresponding author: [adedeji@fuwukari.edu.ng](mailto:adedeji@fuwukari.edu.ng)

### Abstract

This study evaluated quality of fermented maize and moringa seed protein concentrate (MSPC) flour blends. Moringa protein concentrate (10%, w/w) was incorporated at different stages (souring, pre-drying and post drying) during the production of fermented maize flour (FMF). Proximate composition, functional, pasting, colour order and sensory properties of blends were determined using standard methods. Protein, fat and ash contents were 7.86-33.33, 0.14-1.20 and 0.41-1.44%, respectively. There was no significant ( $p>0.05$ ) difference among the samples in terms of energy value. There was a significant ( $p<0.05$ ) reduction in packed density, swelling capacity, peak, breakdown, final and setback viscosity while oil and water absorption capacity, and pasting temperature increased consequent to MSPC inclusion in FMF. Furthermore, there was a significant reduction in  $L^*$  while  $a^*$  and  $b^*$  increased. Combined souring of maize mash and MSPC resulted in improved colour purity as indicated by high delta chrome and colour intensity. In addition, blend produced by this method compared significantly ( $p<0.05$ ) with 100% FMF in terms of colour, consistency, appearance and overall acceptability. This study showed that stage of MSPC inclusion significantly ( $p<0.05$ ) affected the quality of FMF.

**Keywords:** Fermentation; Maize flour; Moringa seed; Protein concentrate

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### 1. INTRODUCTION

Maize (*Zea mays*) is one of the major staple foods in many parts of Western and Central Africa, where it is consumed by no less than 50% of the entire population of the regions (Beugre et al., 2014). According to International Institute of Tropical Agriculture (IITA), annual global production output of maize is estimated at 785 million tons and Africa contributes 6.5% of the total output (IITA, 2019). Nigeria is the top producer in Africa with an average annual production volume of 8 million tons (IITA, 2019).

Due to its high functionality, maize and its products are enjoying increasing applications in many food and non-food systems (Abiose and Ikujenlola, 2014). In West Africa, maize is subjected to different processing operations, and thus, many products are obtained (Nsto et al., 2016). Particularly, production of *ogi*, a popular complementary/breakfast product in

Nigeria, Cameroun and Benin is achieved through submerged fermentation of maize grains (Ayo-Omogie and Ogunsakin, 2013).

*Ogi* serves as a cheap and affordable food for the teeming West African population, however, it is characterised with low nutrient density (Nsto et al., 2016). Approaches for combating malnutrition, such as exogenous and endogenous fortification, provide promising evidence for dramatically improving the health and nutrition of many consumers of the product. Recent works on fortification of *ogi* to improve its nutritional density include supplementation with moringa leaves (Olorode et al., 2013), soybean (Akanbi et al., 2010) and bambara seed (Mbata et al., 2009). Quality improvement of *ogi* through co-fermentation of maize and protein-rich vegetables such as legumes are also well documented (Oyarekua, 2012; Oyarekua, 2013a; Oyarekua, 2013b).

Nutritional improvement of *ogi* with legumes is often characterised with poor acceptability by

consumers owing to impartation of beany flavour (Mume et al., 2013), and reduced digestibility as a result of increased concentration of recalcitrant oligosaccharides in the product (Gayol et al., 2013). Concentration or isolation of protein from legumes prior to incorporation into foods has been reported to reduce these defects (Tang and Liu, 2017; Akinwale et al., 2017). Recent studies showed improvement in chemical composition, physicochemical and sensory properties of cereal-based cookies (Adeyeye et al., 2017; Tang and Liu, 2017) and pasta (Surasani et al., 2019) supplemented with vegetable protein concentrates/isolates. However, there is paucity of information on application of vegetable protein concentrates in fermented cereal-based products. Furthermore, information on stage of vegetable protein concentrate inclusion on quality parameters of foods is sparse. The objective of this study was to determine effects of stage of moringa seed protein concentrate (MSPC) inclusion on proximate composition, functional and sensory properties of fermented maize flour (FMF).

## 2. MATERIALS AND METHODS

### Materials

Moringa seeds and maize grains were sourced from Agbekoya market, Apata, Ibadan and IITA, Ibadan, respectively. Moringa seeds and maize grains were stored in separate sterile containers at refrigeration temperature ( $4\pm 2^{\circ}\text{C}$ ) until required.

### Methods

#### Production of Defatted Moringa Seed Protein Concentrate

Moringa seed flour was produced based on the methodology described by Ijarotimi et al. (2013). Having cleaned the seeds of extraneous materials, they were pretreated with 200 ppm 5.25% sodium hypochlorite for 5 minutes. The pretreated seeds were rinsed in de-ionized water (1:3 w/v) for 9 h at ambient temperature ( $28\pm 2^{\circ}\text{C}$ ), dehulled manually (with the aid of wooden mortar and pestle), and dried in a cabinet drier (Plus11 Sanyo Gallenkamp PLC,

UK) at  $50^{\circ}\text{C}$  for 30 h. Subsequently, the dried seeds were milled in an attrition mill (Glope p44 China), sieved using a 0.425 mm sieve to obtain moringa seed flour. Moringa seed flour was defatted using cold extraction principle outlined by Kudre et al. (2013) using solvent that comprised of chloroform and methanol (2:1 v/v). Moringa seed flour (1:10, w/v) was soaked in the solvent and the mixture was heated in a water bath (NL42OS, England) at  $60^{\circ}\text{C}$  with continuous shaking for 30 minutes. The mixture was rapidly cooled in an ice bath to  $25^{\circ}\text{C}$ . The mixture was centrifuged at  $10,000 \times g$  for 30 minutes at  $10^{\circ}\text{C}$ , air dried at ambient temperature ( $28\pm 2^{\circ}\text{C}$ ) and milled to particle size  $\leq 0.250$  mm.

To obtain moringa seed protein concentrate (MSPC), defatted moringa seed flour was soaked in distilled water (1:10, w/v) and the mixture was agitated in an orbiter shaker (KS 260 Basic, USA) for 45 minutes. Subsequently, pH of the mixture was adjusted to 4.5 with the aid of 4M HCl and centrifuged at  $4000 \times g$  for 20 minutes. After centrifugation, the supernatant was discarded and the procedure was repeated two more times. The resulting precipitate was freeze-dried (BK-FD20-100 Series, USA), packaged and stored ( $-18^{\circ}\text{C}$ ) until required (Gayol et al., 2013).

#### Production of Fermented Maize Flour

Fermented maize flour (FMF) was produced according to procedure described by Oluwamukomi et al. (2005). Wholesome maize grains were steeped in potable water (1:6 w/v) for 48 h. The grains were wet-milled in an attrition mill (globe p44, China) and the resulting slurry was sieved using 0.425 mm sieve. The overtail was discarded while the filtrate was left to settle and sour simultaneously in water for 24 h. After souring, the slurry was sieved with the aid of 0.125 mm sieve. The resulting cake was dried in a cabinet drier (Plus11 Sanyo Gallenkamp PLC, UK) at  $60^{\circ}\text{C}$  for 24 h, milled, sieved (0.250 mm sieve), packaged in a plastic container (ZipLock, China), labeled and stored ( $-18^{\circ}\text{C}$ ) for subsequent analyses.

### Production of Fermented Maize and Moringa Protein Concentrate Flour Blends

Three blends were prepared by incorporating MSPC (10%, w/w) at different identified stages during production of FMF. These stages included souring, pre-drying and post drying. The first blend was prepared by incorporating MSPC into milled maize slurry prior to souring (souring stage). The resulting mixture was thoroughly mixed, soured for 24 h, sieved with the aid of 0.125 mm sieve. The resulting cake was dried in a cabinet drier (Plus11 Sanyo Gallenkamp PLC, UK) at 60°C for 24 h, milled, sieved (0.250 mm sieve), packaged in a plastic container (ZipLock, China), labeled (FBFM) and stored (-18°C) for subsequent analyses.

The second blend was prepared by incorporating MSPC into soured maize slurry prior to drying (pre-drying stage). The mixture was thoroughly mixed and dried in a cabinet drier (Plus11 Sanyo Gallenkamp PLC, UK) at 60°C for 24 h, milled, sieved (0.250 mm sieve), packaged in a plastic container (ZipLock, China), labeled (SBFM) and stored (-18°C) for subsequent analyses.

The third blend was prepared by mixing MSPC and FMF (post-drying stage). The mixture was thoroughly mixed, packaged in a plastic container (ZipLock, China), labeled (TBFM) and stored (-18°C) for subsequent analyses.

### Analyses

#### Proximate, Functional, Pasting and Colour Order Analyses

Protein, fat, ash, moisture, carbohydrate content and energy value of samples were determined using standard methods (AOAC, 2005). Procedures described by Narayana and Narsinga (1992), Sasulki et al. (1996), and Akpata and Miachi (2001) were used for determination of bulk density (loose and packed), water and oil absorption capacity (WAC and OAC) and

swelling capacity, respectively. Pasting profile of samples was evaluated with the aid of a Brabender visco-amylograph (Newport Scientific Pty Ltd. Warriewood NSW, Australia) based on the procedure outlined by Chinma et al. (2013) (2013). Colour order parameters of samples were determined using a handheld colorimeter (Chromameter CR-400/410).

#### Sensory Analysis

Each sample (30%, w/v) was reconstituted in water, heated to 85°C in a water bath (Polyscience WB20L11B) and held at this temperature for 15 minutes. Gruels with pap-like consistency were obtained. The gruels were subjected to sensory analysis using a 9 point hedonic scale where 0 and 9 represented dislike extremely and like extremely, respectively. Fifty panelists comprising of nursing mothers, male and female adults that were familiar with *ogi* were recruited for the analysis. Coded samples were served to the panelists in individual booths under fluorescent light. Samples were rated by panelists for colour, taste, aroma, consistency, appearance, mouthfeel and overall acceptability (Ijarotimi and Keshinro, 2012).

#### Statistical Analysis

All experiments were performed in triplicate. Raw data obtained were subjected to one-way analysis of variance to obtain means and standard deviations at 5% probability level, while Duncan's multiple range test was used to separate the means. These were achieved using Statistical Package for the Social Scientists (SPSS) version 17.0.

### 3. RESULTS AND DISCUSSION

Proximate composition of FMF as influenced by stage of MSPC inclusion is presented in Table 1.

**Table 1 Proximate composition of fermented maize and moringa seed protein concentrate flour blends**

Stage of MSPC (10 g/100g) inclusion in FMF	Protein (%)	Fat (%)	Carbohydrate (%)	Moisture (%)	Ash (%)	Energy value (kCal/100g)
Souring	27.15 <sup>b</sup> ±0.80	1.20 <sup>a</sup> ±0.20	60.89 <sup>b</sup> ±1.09	9.56 <sup>b</sup> ±0.10	1.20 <sup>ab</sup> ±0.87	362.96 <sup>a</sup> ±10.50
Pre-drying	33.33 <sup>a</sup> ±0.69	0.60 <sup>b</sup> ±0.02	54.05 <sup>c</sup> ±2.81	10.62 <sup>ab</sup> ±0.02	1.40 <sup>a</sup> ±0.52	354.92 <sup>a</sup> ±11.06
Post-drying	29.33 <sup>ab</sup> ±1.14	1.10 <sup>a</sup> ±0.10	57.14 <sup>bc</sup> ±2.84	10.99 <sup>a</sup> ±0.08	1.44 <sup>a</sup> ±0.05	355.78 <sup>a</sup> ±5.51
Control (100% FMF)	7.86 <sup>c</sup> ±0.53	0.14 <sup>c</sup> ±0.11	80.65 <sup>a</sup> ±2.99	10.94 <sup>a</sup> ±0.07	0.41 <sup>b</sup> ±0.07	355.30 <sup>a</sup> ±10.24

Values are means±standard deviations of triplicate scores. Means with different superscripts in the same column were significantly different (p<0.05). MSPC Moringa seed protein concentrate), FMF Fermented maize flour.

Protein content of samples was 13.86-37.15% and varied significantly ( $p < 0.05$ ) among the samples. Lowest protein content (13.86%) was recorded for 100% FMF. This implies that inclusion of MSPC in FMF increased protein content significantly ( $p < 0.05$ ). Akanbi et al. (2010) also reported an increment in protein content of *ogi* as a result of addition of soybean flour. Among the blends, FMF that contained MSPC introduced during souring (FBFM) had lowest protein content of 27.15%. This could probably be due to leaching of protein in MSPC during souring and sieving operations. Inyang, and Effiong (2016) had reported loss of nutrients during soaking and sieving operations of fermented cereal products. Fat content of FMF increased significantly ( $p < 0.05$ ) as a result of addition of MSPC. This could be due to influence of traceable fat in MSPC. Furthermore, significantly ( $p < 0.05$ ) higher fat content was recorded in FBFM and FMF that contained MSPC introduced after drying (TBFM) compared to FMF that contained MSPC introduced before drying (SBFM). Carbohydrate content ranged between 54.05 (SBFM) and 80.65% (100% FMF). Inclusion of MSPC caused a significant ( $p < 0.05$ ) reduction in carbohydrate content of FMF. Since carbohydrate content was obtained by difference (AOAC, 2005), its reduction could be attributed to increase in protein content of FMF as a result of MSPC inclusion. Ocheme et al. (2018) also reported reduction in carbohydrate content of wheat flour following supplementation with groundnut protein concentrate. Significantly ( $p < 0.05$ ) lower carbohydrate content obtained for SBFM compared to other blends might be due to its

high protein content. Moisture content was 9.56-10.99%. This was less than 12% threshold considered to be appropriate for flour and stored grains (Iwe et al., 2016). This is advantageous because reduction of moisture content of FMF will reduce proliferation of spoilage organisms especially mould, thus, improving its shelf stability. Ash content ranged between 0.41 (100% FMF) and 1.44% (TBFM). Supplementation of FMF with MSPC resulted in an increase of ash content. This corroborated findings of Ade et al. (2012) who reported an increase in ash content of baked products produced from blend of wheat and water-extractable protein from African yam bean. The increase in ash content could be attributed to combination effect of ash content from FMF and MSPC. Furthermore, significantly ( $p < 0.05$ ) higher ash content was obtained for TBFM (1.44%) and SBFM (1.40%) compared to 1.20% obtained for FBFM. This difference could be connected with loss of water-soluble minerals in MSPC during souring and sieving (Bolaji et al., 2014). This was in contrast with increase in ash content of *ogi* obtained through co-fermentation of maize and soybean (Oluwamukomi et al., 2005). There was no significant ( $p > 0.05$ ) difference among the samples in terms of energy value. This implies that neither supplementation with MSPC nor stage of its inclusion had significant ( $p < 0.05$ ) effect on energy content of FMF.

#### Effect of Moringa Seed Protein Concentrate Inclusion on Functional Properties of Fermented Maize Flour

Table 2 shows functional properties of 100% FMF, and blends of FMF and MSPC.

**Table 2 Functional Properties of fermented maize and moringa seed protein concentrate blends**

Stage of MSPC (10 g/100g) inclusion in FMF	Loose density (g/cm <sup>3</sup> )	Packed density (g/cm <sup>3</sup> )	Oil absorption capacity (mL/g)	Water absorption capacity (mL/g)	Swelling capacity (g/g)
Souring	0.38 <sup>b</sup> ±0.01	0.66 <sup>a</sup> ±0.01	1.41 <sup>c</sup> ±0.05	1.30 <sup>a</sup> ±0.02	6.42 <sup>b</sup> ±0.01
Pre-drying	0.35 <sup>b</sup> ±0.15	0.64 <sup>a</sup> ±0.75	1.90 <sup>a</sup> ±0.06	1.23 <sup>b</sup> ±0.06	5.78 <sup>c</sup> ±0.21
Post-drying	0.37 <sup>b</sup> ±0.02	0.63 <sup>a</sup> ±0.04	1.79 <sup>b</sup> ±0.05	1.19 <sup>c</sup> ±0.09	5.81 <sup>c</sup> ±0.07
Control (100% FMF)	0.52 <sup>a</sup> ±0.01	0.57 <sup>c</sup> ±0.02	1.40 <sup>c</sup> ±0.80	1.10 <sup>d</sup> ±0.01	6.99 <sup>a</sup> ±0.14

Values are means±standard deviations of triplicate scores. Means with different superscripts in the same column were significantly different ( $p < 0.05$ ). MSPC Moringa seed protein concentrate, FMF Fermented maize flour.

Loose and packed bulk density were 0.32-0.38 and 0.57-0.66 g/cm<sup>3</sup>, respectively. Supplementation of FMF with MSPC caused a significant ( $p < 0.05$ ) reduction in loose and packed density. This agreed with reduction in bulk density of maize flour consequent to supplementation with soy protein isolate (Adeyeye et al., 2017).

Reduction in bulk density could be attributed to low carbohydrate content of blends of FMF and MSPC. Complex carbohydrates are often associated with high bulkiness, and supplementation of cereal-based product with vegetable protein reduces its carbohydrate content, and hence, its bulkiness (Gernah et al., 2011). Low bulk density is an advantage because it improves digestibility of food and bioavailability of nutrients (Asaam et al., 2018). Findings in this work showed that loose and packed bulk density of blends of FMF and MSPC were not significantly ( $p > 0.05$ ) different. This shows stability of MSPC irrespective of its stage of inclusion in FMF. This findings contradicted the report of Oyarekua (2012) who reported significant difference in bulkiness of FMF and co-fermented maize and cowpea flour.

Oil absorption and water absorption capacity were 1.40-1.90 and 1.10-1.30 mL/g, respectively. Oil absorption capacity varied significantly ( $p < 0.05$ ) among the blends, however, there was no significant ( $p > 0.05$ ) difference in OAC of 100% FMF and FBFM. High OAC (1.90 mL/g) was obtained for SBFM while low value (1.41 mL/g) was obtained for FBFM. This implied that souring of maize mash and MSPC together resulted in reduction in OAC. This reduction could be

attributed to conformational imbalance between hydrophilic and hydrophobic constituents of the blend during souring operation (Apotiola, 2013). High OAC in SBFM is advantageous since high OAC contributes to organoleptic attributes of foods (Iwe et al., 2016). Significantly ( $p < 0.05$ ) higher WAC was obtained for FBFM compared to SBFM and TBFM. This could be due to modification of starch granules as a result of souring of FMF and MSPC. Adegunwa et al. (2011) had attributed improved WAC to amylose/amylopectin ratio and molecular modification of starch molecules.

Swelling capacity ranged from 5.78 (SBFM) and 6.99 (100% FMF). This implied reduction in swelling capacity of FMF consequent to MSPC supplementation. There was no significant ( $p > 0.05$ ) difference in swelling capacity of SBFM and TBFM. However, swelling capacity obtained for these two samples were significantly ( $p < 0.05$ ) lower than the one obtained for FBFM. High swelling capacity obtained for FBFM could be due to increased hydration of starch consequent to improved activities of lactic acid bacteria during souring operation of maize mash and MSPC. Ocheme et al. (2010) also attributed increased swelling capacity of millet flour to hydration of starch molecules during soaking operation of millet grains.

#### Effect of Moringa Seed Protein Concentrate Inclusion on Pasting Properties of Fermented Maize Flour

Pasting properties of fermented maize and MSPC flour blends are presented in Table 3.

**Table 3 Pasting Properties of fermented maize and moringa seed protein concentrate blends**

Stage of MSPC (10g/100g) inclusion in FMF	Peak V. (mPa.s)	Trough V. (mPa.s)	Breakdown V. (mPa.s)	Final V. (mPa.s)	Setback V. (mPa.s)	Peak time (Min.)	Pasting Temp. (°C)
Souring	1261.67 <sup>d</sup> ±184.75	784.67±108.54	477.00 <sup>d</sup> ±76.21	1220.67 <sup>d</sup> ±176.09	436.00 <sup>c</sup> ±67.55	5.04 <sup>b</sup> ±0.08	78.23 <sup>b</sup> ±0.03
Pre-drying	1683.33 <sup>c</sup> ±81.98	1055.00 <sup>c</sup> ±39.84	628.33 <sup>c</sup> ±42.15	1995.67 <sup>c</sup> ±252.30	940.67 <sup>b</sup> ±12.46	5.44 <sup>a</sup> ±0.08	79.05 <sup>a</sup> ±0.09
Post-drying	3025.67 <sup>b</sup> ±501.14	1724.27 <sup>b</sup> ±279.44	1301.00 <sup>b</sup> ±221.70	3905.33 <sup>b</sup> ±764.99	2180.67 <sup>a</sup> ±85.55	5.38 <sup>a</sup> ±0.04	78.20 <sup>b</sup> ±0.09
Control (100% FMF)	4660.67 <sup>a</sup> ±160.50	2940.00 <sup>a</sup> ±180.13	1720.67 <sup>a</sup> ±19.63	5451.67 <sup>a</sup> ±184.75	2511.67 <sup>a</sup> ±4.62	5.09 <sup>b</sup> ±0.04	76.65 <sup>c</sup> ±0.09

Values are means±standard deviations of triplicate scores. Means with different superscripts in the same column were significantly different ( $p < 0.05$ ). MSPC Moringa seed protein concentrate, FMF Fermented maize flour, V. Viscosity, Temp. Temperature.

Peak, trough, breakdown, final and setback viscosity were 1261.67-4660.67, 784.67-29040.00, 477.00-1720, 1220.67-5451.67 and 436.00-2511.67 mPa.s, respectively. Findings in this study showed significant ( $p < 0.05$ ) reduction of the listed pasting parameters consequent to inclusion of MSPC. This implied interference of FMF starch swelling due to presence of MSPC (Asaam et al., 2018). Akinwale et al. (2017) also reported reduction in viscosity of cassava starch-based custard following inclusion with soy protein concentrate. High pasting profile recorded for 100% FMF could be attributed to its high carbohydrate content (Section 3.1) (Fila et al., 2013). Also, stage of MSPC inclusion had significant ( $p < 0.05$ ) effect on the pasting parameters. Results showed that lowest and highest peak, trough, breakdown, final and setback viscosity were recorded for FBFM and TBFM. Low pasting profile recorded for FBFM could be due to denaturation of protein during fermentation and consequent reduction in hydrophilic tendency of denatured proteins (Oluwamukomi et al., 2005). Differences in peak viscosity of the blends could be attributed to differences in rate of starch granule swelling and water absorption (Asaam et al., 2018).

Peak time varied between 5.04 (FBFM) and 5.44 minutes (SBFM). There was no significant ( $p > 0.05$ ) difference between FBFM and 100% FMF in terms of peak time. Low peak time of FBFM is an indication of short preparation time of gruel which is a good advantage (Chinma et al., 2013). Generally, low pasting temperature (76.65-79.05°C) was recorded for the samples. These values were lower than 81.10-89.80°C reported for blends of maize-African yam bean flour (Idowu, 2015).

Inclusion of MSPC in FMF caused a significant ( $p < 0.05$ ) increase in pasting temperature. This contradicted findings of Akinwale et al. (2017) who reported that inclusion of soy protein isolate did not cause any significant change on pasting temperature of cassava starch-based custard. Significantly ( $p < 0.05$ ) lower pasting temperature was recorded for FBFM and TBFM compared to SBFM. This implied lower energy requirement during preparation.

#### Effect of Moringa Seed Protein Concentrate Inclusion on Colour Order Properties of Fermented Maize Flour

Table 4 shows colour order parameters of fermented maize flour and MSPC blends.  $L^*$  (measure of lightness) ranged from 81.31 (FBFM) to 93.11 (100% FMF). Results showed that addition of MSPC into FMF caused a significant ( $p < 0.05$ ) reduction in  $L^*$ . Colour of MSPC was creamy white and its inclusion in FMF might have caused reduction in  $L^*$  of FMF. This result corroborated the findings of Tang and Liu (2017) who reported reduction in  $L^*$  of cookie as a result of partial replacement of wheat flour with whey and soy proteins. In addition,  $L^*$  varied significantly ( $p < 0.05$ ) among blends of FMF and MSPC with FBFM and TBFM having lowest and highest value, respectively. Low  $L^*$  recorded for FBFM could be due to leaching of pigments from MSPC during souring operation and subsequent absorption of leached pigments by FMF during sieving operation (Ojinnaka et al., 2013). Degree of redness ( $a^*$ ) and yellowness ( $b^*$ ) of samples were 0.14-0.75 and 7.79-15.30, respectively. Both parameters decreased significantly ( $p < 0.05$ ) consequent to MSPC inclusion.

**Table 4 Colour order parameters of fermented maize flour and moringa seed protein concentrate blends**

Stage of MSPC Inclusion in FMF	L	$a^*$	$b^*$	Hue angle	Delta chrome	Colour intensity
Souring	81.31 <sup>d</sup> ±4.92	0.75 <sup>a</sup> ±0.13	15.30 <sup>a</sup> ±0.58	87.20 <sup>b</sup> ±0.39	14.99 <sup>a</sup> ±0.57	16.28 <sup>a</sup> ±1.67
Pre-dering	88.70 <sup>c</sup> ±5.10	0.41 <sup>b</sup> ±0.15	13.58 <sup>b</sup> ±0.67	88.28 <sup>a</sup> ±0.59	13.26 <sup>b</sup> ±0.67	14.02 <sup>b</sup> ±1.76
Post drying	90.76 <sup>b</sup> ±3.57	0.46 <sup>b</sup> ±0.13	8.64 <sup>c</sup> ±1.19	86.95 <sup>c</sup> ±0.92	8.32 <sup>c</sup> ±1.19	9.88 <sup>c</sup> ±1.60
Control (100% FMF)	93.11 <sup>a</sup> ±4.24	0.14 <sup>c</sup> ±0.12	7.79 <sup>d</sup> ±1.73	87.67 <sup>b</sup> ±1.23	7.47 <sup>c</sup> ±1.72	10.24 <sup>c</sup> ±3.74

Values are means±standard deviations of triplicate scores. Means with different superscripts in the same column were significantly different ( $p < 0.05$ ). MSPC Moringa seed protein concentrate, FMF Fermented maize flour.

**Table 5 Sensory properties of gruel produced from fermented maize and moringa seed protein concentrate blends**

Stage of MSPC FMF	Colour	Taste	Aroma	Consistency	Appearance	Mouthfeel	Overall acceptability inclusion in
Souring	8.18 <sup>a</sup> ±1.35	7.34 <sup>b</sup> ±1.26	7.54 <sup>b</sup> ±1.50	7.06 <sup>a</sup> ±1.63	8.02 <sup>a</sup> ±1.32	6.96 <sup>b</sup> ±1.59	7.44 <sup>a</sup> ±1.31
Pre-drying	7.30 <sup>b</sup> ±0.93	5.74 <sup>c</sup> ±1.31	6.52 <sup>b</sup> ±1.37	5.96 <sup>b</sup> ±1.11	6.42 <sup>b</sup> ±1.26	6.48 <sup>c</sup> ±1.20	6.64 <sup>bc</sup> ±1.05
Post-drying	5.38 <sup>c</sup> ±1.96	5.78 <sup>d</sup> ±1.15	5.84 <sup>c</sup> ±1.25	5.72 <sup>b</sup> ±1.40	6.00 <sup>c</sup> ±1.46	5.48 <sup>d</sup> ±1.34	6.46 <sup>c</sup> ±1.40
Control (100% FMF)	8.26 <sup>a</sup> ±1.37	8.20 <sup>a</sup> ±2.21	7.94 <sup>a</sup> ±1.48	6.98 <sup>a</sup> ±1.76	8.20 <sup>a</sup> ±1.47	7.40 <sup>a</sup> ±1.34	7.60 <sup>a</sup> ±1.09

Values are means±standard deviations of 50 scores. Means with different superscripts in the same column were significantly different ( $p < 0.05$ ). MSPC Moringa seed protein concentrate, FMF Fermented maize flour.

This implied increased colouration and deviation from white colour (Ojinnaka et al., 2013). Higher  $b^*$  was recorded for the blends as compared to  $a^*$  and this implied higher deviation towards yellow than red. This could also be due to white creamy colouration of MSPC. Similar result was obtained for co-processed quality protein maize and carrot complementary food (Oladeji, 2018).

Hue angle, delta chrome and colour intensity were 86.95-88.28, 7.47-14.99 and 9.88-16.28, respectively. Hue angle obtained in this study showed deviation between 0 and 90° and this implied deviation from red to yellow. This agreed with with results obtained for  $a^*$  and  $b^*$ . Significantly ( $p < 0.05$ ) higher delta chrome and colour intensity were obtained for FBFM and SBFM compared to TBFM and 100% FMF. This indicated darker colouration of these blends probably as a result to synthesis of pigments during co-souring and drying of maize mash and MSPC. Similar result was obtained for wheat-soy protein isolate-based cookie (Tang and Liu, 2017). High chroma and colour intensity recorded for FBFM and SBFM are indication of high colour purity of the blends (Correia et al., 2016).

#### **Sensory Properties of Gruel Produced from Fermented Maize and Moringa Seed Protein Concentrate Flour Blends**

Sensory properties of samples in terms of colour, taste, aroma, consistency, appearance, mouthfeel and overall acceptability are presented in Table 5. There was no significant ( $p > 0.05$ ) difference between FBFM and 100% FMF in terms of colour, consistency,

appearance and overall acceptability and their scores were significantly ( $p < 0.05$ ) higher than those of SBFM and TBFM. In addition, higher scores were obtained for FBFM compared to SBFM and TBFM in terms of taste, aroma and mouthfeel. This implied better consumer acceptability of FBFM than other blends. Low acceptability recorded for SBFM and TBFM agreed with the findings of Tang and Liu (2017) who reported reduction in consumer acceptability of cookie with increasing level of whey and soy proteins. High consumer preference of FBFM showed the importance of co-souring of maize and MSPC and this conformed with the finding of Oyarekua (2013b) who reported improved sensory properties of a complementary food as a result of co-fermentation of maize, cowpea and sweet potato.

#### **4. CONCLUSION**

This study showed that inclusion of MSPC improved proximate composition, functional, pasting, colour order and sensory properties of FMF. Also, these parameters were significantly ( $p < 0.05$ ) affected by stage of MSPC inclusion in FMF. Inclusion of MSPC in FMF resulted in an increase in protein, fat and ash while carbohydrate reduced. Co-souring of maize mash and MSPC resulted in reduction of protein and ash contents. FBFM compared significantly with 100% FMF in terms of colour, consistency, appearance and overall acceptability.

## Declaration of Conflict of Interest

None declared

## Authors' Contributions

OOE designed the study and approved the final draft and OEA conducted laboratory experiment, statistical analysis and prepared the first draft.

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